Association analysis of C677T and A1298C polymorphisms in MTHFR gene in patients with colorectal cancer susceptibility

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Highlights

• 5,10-methylenetetrahydrofolate gene may cause impairment of cell function and a variety of cancers like prostate and colorectal cancers.
• MTHFR C677T and A1298C polymorphism can increase the susceptibility risk of colorectal cancer.
• There are significant associations between genotypes 677CT and 677TT with colorectal cancer risk.
• Inflammatory mechanisms are associated with the disorders of female reproductive system.

Graphical Abstract

Abstract

5,10-methylenetetrahydrofolate reductase (MTHFR) plays an important role in folate metabolism. Also, this gene is associated with repair, synthesis and methylation of DNA. Polymorphisms of C677T and A1298C are two common SNP of MTHFR gene. This study was aimed to investigate the association of C677T and A1298C polymorphisms in MTHFR with colorectal cancer in the Iranian population. In this case-control study, 100 patients with colorectal cancer and 100 healthy individuals' samples as a control group were collected. Genomic DNA was isolated from peripheral blood samples, and MTHFR C677T and A1298C genotyping were performed via RFLP-PCR. Significant associations among genotypes 677CT (OR= 2.0910, 95% CI= 1.1728 to 3.7280, p= 0.0124) and 677TT (OR= 5.2073, 95% CI= 1.0299 to 26.3283, p= 0.0460) with risk of colorectal cancer was found. But no significant relationship was detected between A1298C polymorphism and colorectal cancer. MTHFR C677T polymorphism can increase the risk of colorectal cancer in the Iranian population.

Keywords:
Colorectal cancer
Methylenetetrahydrofolate
Genetic polymorphism
PCR-RFLP

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Introduction

Colorectal cancer (CRC) is the third most common cancer in men and the fourth leading cause of cancer death in the world. In addition, it is expected to increase by 60% and 2.2 million new and 1.1 million deaths by 2030 (1). Some risk factors including age, tobacco usage, consumption of red meat, family history and genetic factors are involved in colorectal cancer (2). A recent study determined folate, a water-soluble vitamin, plays an important role in colorectal cancer (3). Deficiency in the synthesis of folic acid results in failure in the double-strand chromosome that it is a methyl transporter in the metabolism of the cell and its deficiency leads to genomic hypomethylation and ultimately disrupts the expression of the genes.

The human 5, 10-methylenetetrahydrofolate reductase (MTHFR) gene is located on chromosome 1 (1p36.3) and composed of 12 exons, encompasses 19.3 kb of DNA and the gene codes a 74.6-kDa protein of 656 amino acids (4). 5, 10-methylenetetrahydrofolate reductase gene plays an important role in the metabolism of folate. MTHFR catalyzes the conversion of 5, 10-methylenetetrahydrofolate (5,10-MTHF) to 5- methyltetrahydrofolate (5-MTHF) which is the main form in the circulation of folate in the blood and lead to the conversion of dUMP (deoxyuridine monophosphate) to dTMP (deoxythymidine monophosphate). Also, MTHFR protein causes biosynthesis of DNA, RNA and homocysteine re-methylation. Considering the important regulatory role of MTHFR on DNA replication, DNA repair, DNA methylation and cell division, this gene can have the potential role for cancer-predisposing and consequently low dietary folate and MTHFR deficiency causing to intestinal tumors in the BALB/c mouse model (5). Low levels of 5, 10-MTHF can lead to an increased dUMP/dTMP ratio, increased risk of DNA mutation and the fracture of DNA strand (6). Also, low levels of 5, 10-MTHF can lead to decreased s-adenosylmethionine (SAM) levels and DNA hypomethylation can cause to genomic instability, activation of cellular oncogenes and DNA damage. However, Low levels of folate can be associated with susceptibility to cancer (7). There are three common polymorphisms of the MTHFR gene, MTHFR C677T (rs1801133), G1793A (rs2274976) and MTHFR A1298C (rs1801131). In the case of the MTHFR C677T polymorphism, the base of cytosine at position 677 changes to a thymidine base while this change leads to conversion in the amino acid at position 222 so that alanine is replaced by valine on the other hand in the 1298A>C transition results from a glutamate to alanine replacement at codon 429 (Glu429Ala) and eventually the 1793G>A polymorphism leads to in an arginine-to-glutamine replacement at position 594 (Arg594Gln). MTHFR C677T can be as a risk factor for heart disease, congenital anomalies, diabetes, stroke, psychiatric disorders and some types of cancer (8). Considering the role of MTHFR in impaired DNA methylation, DNA strand breaks and repair and relationship between MTHFR with folate deficiency and colorectal cancer the present study, the association of C677T and A1298C polymorphisms in MTHFR with colorectal cancer in an Iranian population, has been analyzed.

Materials and Methods

Collection and analysis of blood samples

This study performed on 100 patients with colorectal cancer and 100 healthy individuals’ samples as a control group. Peripheral blood samples were collected from Isfahan University of Medical Sciences, Isfahan, Iran into tubes containing anticoagulant sodium citrate and preserved at -20 °C for further usage.

Genomic DNA extraction and primers

Isolation of genomic DNA was done using blood samples and Whole DNA of blood samples was isolated using DNA extraction kit (Add bio Co) and kept at -20°C until usage. The polymerase chain reaction was performed to amplify the MTHFR fragments, in 25 µL PCR reaction containing 0.35 µM forward and reverse primers, 0.5 µL dNTPs mix, 0.2 µL Taq polymerase, 35 ng template DNA and 1.5 µM MgCl2 (All PCR reagents were purchased from Fermentas) (Figure 1).
Figure 1. PCR product fragments on 1% agarose gel stained with 1 mg/mL ethidium bromide solution. L, DNA marker, 233-bp fragment of the 677C.T transition, 143-bp fragment of the 1298A.C transition.

Primer sequences for two single nucleotide polymorphisms (SNPs), C677T (rs1801133) and A1298C (rs1801131) are presented in Table 1. The PCR was performed in a thermal cycler (Eppendorf, Hamburg, Germany) using the conditions listed in Table 2. The PCR products were evaluated by 1% agarose gel electrophoresis. Then, 5 mL aliquot of the PCR product was digested with 5 units HinfI and MboII (Fermentas) and being incubated at 37 °C for 16 h and finally digested products were evaluated by polyacrylamide gels and silver nitrate (Figure 2). Results of PCR-RFLP data were checked by DNA direct sequencing.

Table 1. Sequences of the RFLP-PCR Primer.

<table>
<thead>
<tr>
<th>SNP (rs1801133)</th>
<th>Primer</th>
<th>Primer sequence</th>
</tr>
</thead>
<tbody>
<tr>
<td>C677T (rs1801133)</td>
<td>MTHFR1-forwarded</td>
<td>5’-CGAAGCAGGGAGCTTTGAGGCTG</td>
</tr>
<tr>
<td></td>
<td>MTHFR1-reverse</td>
<td>5’-AGGACGGTGCGGTGAGAGTG</td>
</tr>
<tr>
<td>A1298C (rs1801131)</td>
<td>MTHFR2-forwarded</td>
<td>5’-GCAAGTGCCCCCAAGGAGG</td>
</tr>
<tr>
<td></td>
<td>MTHFR2-reverse</td>
<td>5’-GGTCCCCACTCCAGCATC</td>
</tr>
</tbody>
</table>

Table 2. Polymerase chain reaction conditions.

<table>
<thead>
<tr>
<th>SNP (rs1801133)</th>
<th>PCR conditions</th>
<th>PCR product (bp)</th>
</tr>
</thead>
<tbody>
<tr>
<td>C677T (rs1801133)</td>
<td>42 cycles of 94°C for 5 min, 94°C for 45 s, 63°C for 50 s and 72°C for 1 min; final extension at 72°C for 10 min</td>
<td>233</td>
</tr>
<tr>
<td>A1298C (rs1801131)</td>
<td>45 cycles of 94°C for 5 min, 94°C for 45 s, 55°C for 1 min and 72°C for 1 min; final extension at 72°C for 10 min</td>
<td>143</td>
</tr>
</tbody>
</table>

Figure 2. Polyacrylamide gel electrophoresis results of the restriction enzyme map. Restriction enzyme map of the 233-bp polymerase chain reaction (PCR) fragment of the methylenetetrahydrofolatereductase (MTHFR) gene following digestion with HinfI. M, DNA marker; Lane 1, CC genotype; Lane 2, TT genotype; Lane 3, CT genotype.
Statistical analysis

Hardy-Weinberg equilibrium in case and control groups for both C677T and A1298C polymorphisms were evaluated by chi-squared test. The differences of genotype and allele frequencies between case and control groups were also evaluated by the same test. The strength of association between C677T and A1298C polymorphisms and colorectal cancer risk was estimated by odds ratio (OR) and 95% confidence interval (CI) which were computed by binary logistic regression analysis. A P-value less than 0.05 were considered statistically significant. Statistical analysis was performed using SPSS ver.19 software.

Results

The results of genetic association study are presented in Tables 3 and 4. With regard to polymorphism C677T, we found that there are significant associations between genotypes CT (OR= 2.0910, 95%CI= 1.1728 to 3.7280, p= 0.0124) and TT (OR= 5.2073, 95%CI= 1.0299 to 26.3283, p= 0.0460) and colorectal cancer risk. Also, there was a significant association between the mentioned polymorphism and colorectal cancer in a dominant model (OR= 2.2508, 95%CI= 1.2781 to 3.9639, p= 0.0050). In addition, there was a significant association between allele T and risk of colorectal cancer (OR= 1.9101, 95%CI= 1.2149 to 3.0030, p= 0.0051). However, we did not find any significant association between A1298C polymorphism and colorectal cancer. Obtained data revealed that there are no significant associations between genotypes AC (OR= 1.2401, 95%CI= 0.6806 to 2.2596, p= 0.4821) and CC (OR= 0.9790, 95%CI= 0.3840 to 2.4960, p= 0.9646) and colorectal cancer. Moreover, there was no significant association between A1298C and colorectal cancer in a dominant model (OR= 1.1748, 95%CI= 0.6733 to 2.0498, p= 0.5705). Allele analysis revealed that there is no significant association between allele C and colorectal cancer (OR= 1.0768, 95%CI= 0.6967 to 1.6644, p= 0.7390).

Table 3. Genotype frequencies of C677T and A1298C polymorphisms in cases and controls.

<table>
<thead>
<tr>
<th>Polymorphism</th>
<th>Genotype</th>
<th>No. and Percentage</th>
<th>OR (95% CI)</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Control (n=100)</td>
<td>Case (n=100)</td>
<td></td>
</tr>
<tr>
<td>C677T</td>
<td>CC (%)</td>
<td>61 (61.00)</td>
<td>(41 41.00)</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>CT (%)</td>
<td>37 (39.39)</td>
<td>52 (52.00)</td>
<td>2.0910 (1.1728 to 3.7280)</td>
</tr>
<tr>
<td></td>
<td>TT (%)</td>
<td>2 (2.00)</td>
<td>7 (7.00)</td>
<td>5.2073 (1.0299 to 26.3283)</td>
</tr>
<tr>
<td></td>
<td>CT+TT (%)</td>
<td>39 (39.00)</td>
<td>59 (59.00)</td>
<td>2.2508 (1.2781 to 3.9639)</td>
</tr>
<tr>
<td>A1298C</td>
<td>AA (%)</td>
<td>56 (56.00)</td>
<td>52 (52.00)</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>AC (%)</td>
<td>33 (33.00)</td>
<td>38 (38.00)</td>
<td>1.2401 (0.6806 to 2.2596)</td>
</tr>
<tr>
<td></td>
<td>CC (%)</td>
<td>11 (11.11)</td>
<td>10 (10.00)</td>
<td>0.9790 (0.3840 to 2.4960)</td>
</tr>
<tr>
<td></td>
<td>CC+CA (%)</td>
<td>44 (44.00)</td>
<td>48 (48.00)</td>
<td>1.1748 (0.6733 to 2.0498)</td>
</tr>
</tbody>
</table>

*Significant difference between patient and control group.

Table 4. Allele frequencies of C677T and A1298C polymorphisms in cases and controls.

<table>
<thead>
<tr>
<th>Polymorphism</th>
<th>Allele</th>
<th>No. and Percentage</th>
<th>OR (95% CI)</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Control (n=200)</td>
<td>Case (n=200)</td>
<td></td>
</tr>
<tr>
<td>C677T</td>
<td>C (%)</td>
<td>159 (79.50)</td>
<td>134 (67.00)</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>T (%)</td>
<td>41 (20.50)</td>
<td>66 (33.00)</td>
<td>1.9101 (1.2149 to 3.0030)</td>
</tr>
<tr>
<td>A1298C</td>
<td>A (%)</td>
<td>145 (72.50)</td>
<td>142 (71.00)</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>C (%)</td>
<td>55 (27.50)</td>
<td>58 (29.00)</td>
<td>1.0768 (0.6967 to 1.6644)</td>
</tr>
</tbody>
</table>

*Significant difference between patient and control group.
Discussion

Folate is a form of vitamin B9 that it has an important role in the different function. Mammalian cells cannot produce folate and leafy vegetables, liver, potatoes, bread and fruit are important sources of folate. Recent studies provided crucial evidence that folate deficiency is associated with an increased risk of numerous cancer especially in colorectal cancer because of lower intakes of vegetables and fruit (9) Folate plays an important role in modification reactions including DNA methylation, DNA synthesis, nucleotide synthesis and repair (10). The MTHFR gene encodes a cytoplasmic flavor enzyme and regulatory enzyme involved in folate metabolism through the production 5-methyltetrahydro folate that this form dominant circulating form of folate (11, 12). Thus failure and mutations in the MTHFR gene may cause impairment of cell function and a variety of cancers so myeloid leukaemia, prostate cancer and colorectal cancer. The present study aimed to investigate the association between C677T and A1298C polymorphisms in MTHFR with colorectal cancer in an Iranian population. The results of this study showed a significant association between colorectal cancer and C677T genotypes but in the A1298C polymorphism was not associated with the risk of colorectal in this study. Several studies have been conducted to investigate the association between C677T, A1298C polymorphisms and colorectal cancer in different populations which confirmed the results of this study. In several studies, the 677TT genotype was associated with an increased risk of CRC. The results of the Sameer et al., study showed that the MTHFRC677T polymorphism slightly increases the risk for colorectal cancer development in the ethnic Kashmir population that is consistent with the results of this study (13).

Also, the results of a study by Ozen and et al., suggested a strong association between both SNPs of MTHFR 677 C>T and 1298 A>C and CRC susceptibility in the Turkish population (14). In addition, the data of the study Gallegos-Arreola et al., determined that the 677C>T polymorphism in MTHFR increased significantly the risk of CRC in the Mexican population (15). But, the multiethnic study conducted by Le Marchand et al., showed the negative correlation of the TT genotype with colorectal cancer, especially those diagnosed at an advanced stage (16). Furthermore, the research results of Cao et al., showed that polymorphisms of the MTHFR 677 C>T could impress susceptibility to colon cancer and interestingly when women and men were evaluated together there was no significant relationship between MTHFR C677T and A1298C polymorphisms with colon cancer but when the two groups of women and men get separated from each other, different results were obtained, men with MTHFR 677 C>T T/T genotype have a significantly higher risk for colon cancer also a significant association between 677 C>T T/T and 1298 A>C A/A genotypes with colon cancer (17). Single nucleotide polymorphisms may influence the expression and structure of a gene depending on their locations. Polymorphisms in the promoter region could affect the gene expression while the exonic polymorphism could alter the function and structure of a protein (18, 19, 20, 21, 22).

Also, mutations in the intron regions might change the gene expression by interfering in the splicing process. Evaluation of the molecular effects of single nucleotide polymorphisms on the interested genes by in vitro and in vivo methods could be a time and cost consuming procedures. But, the employing of computational methods can be a beneficial method to evaluate the molecular aspects of an SNP. A previous study revealed that the C677T transition has a potential effect on both RNA and protein structure. They present that this mutation may be deleterious for the protein structure due to its location near the substrate-binding site. There are many reports about the effects of genotype on cancer, and more research is needed to show more angles (23, 24, 25, 26).

Conclusion

According to results, the 677 C>T polymorphism MTHFR gene can increase the risk factor of developing colorectal cancer, in addition, individuals with T allele in the 677 C>T MTHFR increased risk of developing colorectal cancer also can 677 C>T polymorphism be identified as a risk factor for colorectal cancer but for 1298 A>C, it is needed to supplementary studies with larger sample size. However, the evaluation of gene-gene and gene-environment interaction has not performed and this could be considered as a limitation of this study.
References


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